



## **Genetics, Biochemistry and Biophysical Analysis of Anthocyanin in Rice (*Oryza sativa L.*)**

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**Abstract.** Rice (*Oryza sativa L.*) is the primary staple food for half of the world population. It is generally classified based on the grain color into black, red, purple, brown, green, and white. These colored rice are determined by the composition and concentration of anthocyanin pigments in different layers of aleurone, pericarp, and seed coat. Anthocyanins are also accumulated in various tissues of the rice plants, mostly in the grain, but are also presents in leaves, leaf sheath, floral organ, and hull. The type and concentration of the anthocyanins in rice tissues are influenced by the cultivars and developmental stages. Anthocyanin-enriched rice is related to the health effects, including antioxidant, antibacterial, and anti-inflammation activities that potentially use as functional food ingredients, dietary supplements, and natural colorants. Structural and regulatory genes are involved in anthocyanin biosynthesis of rice. Various molecular biology techniques have been applied to improve productivity, nutritional contents, and market value of pigmented rice. This review focused on the genetics, biochemistry and biophysical analysis of anthocyanin in rice that will facilitate rice breeding program to develop new high-yield pigmented rice varieties.

**Keywords:** rice, anthocyanins, genetics, biochemistry, biophysics

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### **1. Introduction**

Rice (*Oryza sativa L.*) has become the major staple food for almost half of the global population due to the nutrients composition, including carbohydrate, protein, oil components, and other micronutrients [1-5]. There are various kinds of rice are consumed that can be classified based on the grain color into black, red, purple, brown, green, and white [6-9]. White rice is generally consumed, while pigmented rice, such as black, red, purple, and brown contain natural pigment anthocyanins that accumulate in the particular layers of the seed coat, pericarp, and aleurone [10-18]. Total anthocyanins in each pigmented rice

varieties are diverse in the range of 0-493 mg/100 g [19]. Black rice is the most common pigmented rice in the market due to their sensory characteristics and organoleptic properties, such as good taste, fragrant aroma, fluffiness texture, high nutrition values, and positive health effects [20-27]. China is the biggest black rice producer followed by Sri Lanka, Indonesia, India, Philippines, Bangladesh, Malaysia, Thailand, and Myanmar [17]. Pigmented rice genotypes have been cultivated in Asia for a long time, such as Chinese black rice, Indonesian black rice, and Thai black rice [20]. Anthocyanins of the pigmented rice has the potential to be applied as a dietary supplements, functional food ingredient, and natural colorant for food, beverages, and pharmaceutical products. Anthocyanins show positive effects for the human health, including antioxidant, antibacterial, anti-inflammatory, anticancer, anti-diabetic, antitumor, anti-allergic agents, anticarcinogenic, anti-atherosclerosis, and others [23], [28-37], [38-40]. Consequently, anthocyanins play important roles in preventing human diseases, such as atherosclerosis [12], [28], [41], diabetes [42-44], and cancer [45-49]. In the rice plants, anthocyanins involve in pollination to attract the insects, UV-B protection, hormone responses regulation, photo-perception in autumn leaves, stabilize photosynthetic activity, biotic and abiotic stress defense system [50-59].

Anthocyanins are water-soluble natural pigments classified to the phenolic compounds of flavonoids group which responsible for attractive colors, such as purple, red or brown in different tissues of the rice plants with various concentration and composition of anthocyanins [60-64]. There are six anthocyanin types, including malvidin, peonidin, cyanidin, delphinidin, petunidin, and pelargonidin [65]. Different combinations of anthocyanins lead to different colors, the higher anthocyanins concentration and combination, the blacker the color. The concentration and composition of anthocyanins in the rice plants are vary depend on the rice variety and developmental stages [66-68]. The common anthocyanin types in the pigmented rice are cyanidin-3-glucoside, while the minor is peonidin-3-glucoside [69-70]. Anthocyanins are important secondary metabolites in the rice plants and accumulate in various tissues and organs that related to photosynthesis, reproduction, and defense, such as pericarp, aleurone, awn, leaf blade, leaf sheath, palea, lemma, internode, stigma, apiculus, and root [71-74]. Different level of purple color are identified in these tissues and organs of the rice plants. For example, purple and red rice contain 20 times higher concentrations of the anthocyanin in their aleurone layer compared with brown rice [75]. Black rice contain higher total antioxidant capacity compared to red rice; black rice have 0.5-2.5% while red rice only 0.03-0.1% [76]. The purple pigmentation is regulated by the allelic variation of genes, the co-segregation of the alleles do not always happen [72]. Anthocyanin biosynthesis is controlled by the structural and regulation genes, and the stability of the anthocyanins also influenced by the environmental conditions, including temperature, pH value, lights, enzymes, oxygen, and metallic ions [51], [77-84]. Anthocyanin biosynthesis can be enhanced by environmental modification, such as maintaining temperature ranging from 22 to 27°C and light intensity between 301-600 lx. Maintaining the stability of anthocyanins is a crucial factor in food and pharmaceutical industry [85]. Accumulation of anthocyanins in the rice plants are depend on the rice developmental stages, including seedling, vegetative, reproductive, and mature stages. During maturity stage, black rice contains higher anthocyanins in the aleurone layer compared with grain filling stage [75].

Anthocyanins have been used in human diet for centuries as herbal medicines to cure several health problems, such as cold, diarrhea, and hypertension. Recently, anthocyanins are being applied as natural colorant of food and beverages, and also dietary supplements due to their attractive colors and health benefits. In the United States, estimated anthocyanins consumption is around 12.5 mg/day [86]. Black rice as the common pigmented rice in the market that contain various combinations of anthocyanins, such as cyanidin-3-glucoside, peonidin-3-glucoside, and petunidin-3-glucoside in the aleurone layer of the rice grain [13], [17], [70], [76], [87-88]. Cyanidin-3-glucoside is the most anthocyanins composition in black rice around 631 mg/100 g, while peonidin-3-glucoside is around 363 mg/100 g [13], [70], [88-89]. Southeastern Asian countries are the primary producer of black rice. Recently, California also produces black rice due to high market demand [90]. European countries that cultivate black rice are Italy and Greece [23]. In Asian countries such as Thailand, China, Indonesia, India, Korea, and Japan, black rice is usually combine with white rice to increase the flavor [12], [91-94]. The color of cooked black rice become regal purple [95]. Black rice is usually use as food ingredients in fried rice, paella,

risotto, porridge, bread, pasta, and rice cake [96-97]. Black rice also become an important material to produce alcoholic beverages with red color. There are many varieties of black rice, including short and long grains, glutinous and non-glutinous rice, early and late maturity period [98-100]. Red rice is generally used as a natural colorant in ice cream, bread, and liquor [11], [101]. Pigmented rice also contains higher concentration of micronutrients, such as iron, manganese, and zinc in the grain compared to white rice [102-107]. These pigmented rice has a potential to decrease malnutrition around the world [66]. Many studies have been reported the nutritional values of black and red rice [23], [41], [108-111]. Powdered anthocyanins extracts from black rice was produced with spray-drying and freeze-drying processes of Italian black rice that contain rich anthocyanins and antioxidant activity [112]. These powdered anthocyanins are more stable from environmental conditions of storage and food processing, including temperature, pH value, and lights that give high economic value to use for functional foods and pharmaceuticals products [6], [79], [113-114].

In China, red rice has been approved by the Chinese Ministry of Health as a food colorant in soybean products, meat, and fish [115]. Meanwhile, in United States, Canada, European Union, Japan, Australia, New Zealand, and South Africa restrict anthocyanins as a food colorant [116]. Red rice is commonly use as food colorants and dietary supplements in China due to attractive and high stability color from high light exposure, pH changes, and heat conditions; good taste and flavor; cheap; high availability; and high yield rice variety [117]. The anthocyanin concentration in red rice is lower than black rice around 1.5 to 9.4 mg/100 g [70], [118]. Fermented rice as a dietary supplement to decrease cholesterol accumulation of the blood circulation has been marketed in China. Red rice as a herbal medicine to treat cardiovascular disease and abdominal pain [11].

Analysis of biosynthesis, storage, and transportation mechanisms in anthocyanins have been achieved a significant progress due to the development of molecular biology. These progress give a positive impact on the food industry, pharmaceuticals, flavors, rice breeding program [119-121]. Identification of physicochemical properties, quantification, and extraction of anthocyanins in the rice plants was based on Ultraviolet-Visible (UV-Vis) absorption spectrophotometer, High Performance Liquid Chromatography (HPLC), mass spectrometry, liquid chromatography, and paper chromatography [101], [104], [122-124]. Based on the UV-Vis absorption spectrophotometer data, anthocyanins show maximum absorption range in the region 500-535 nm of the blue spectrum; malvidin at 530 nm, peonidin at 517-520 nm, cyanidin at 512-520 nm, delphinidin at 525 nm, petunidin at 526-529 nm, and pelargonidin at 502-506 nm [70]. By identification of phytochemical properties of anthocyanins in pigmented rice gives insights to the application of pigmented rice as health promotion agents [122], [125-126]. Based on the genetic analysis of pigmentation in pigmented rice, there are three genes that regulates the pigmentation, including Ra, Rc, and Rd genes. The intensity of the pigmented rice coloration is influenced by the presence of genes and the genes status (dominant or recessive). Ra genes regulated purple pericarp, which purple color is dominant and white color is recessive. Brown pericarp is produced when Rc gene presence and Rd gene absence. Both of the genes, Rc and Rd genes are presence produce red pericarp. Meanwhile, if only Rd gene presence, it will not produce any color [99], [127-128]. The alleles segregation of coloration in pigmented rice also presence, for example F2 population of the crossing between black rice and white rice varieties showed three phenotypes; black, brown, and white color with the segregation ratio 9:3:4.

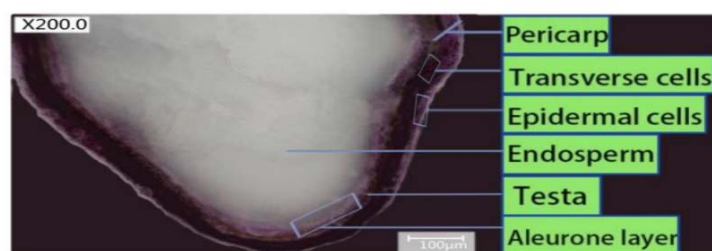
Pigmented rice cultivation since ancient time and in 1970s become more popular due to the development of genetic engineering [129]. The famous pigmented rice varieties in Korea are dark red Heugjinjubyeo, dark purple Heugnabyeo, dark blue Jakwangdo, red brown Sanghaehyeolla, black purple Hongmi, and dark red-purple Kilimheugmi [130]. The quality of pigmented rice varieties have been improve by employing recent technologies, including genome sequencing, gene expression analysis, gene editing, and omics technologies [102], [131-134]. High-yield pigmented rice varieties have been developed by applying the characteristics of grain pigmentation inheritance, tagging the key genes that controlled the rice quality traits, and identifying markers of these rice quality traits [135-136]. The advanced pigmented rice varieties can be developed by understanding the molecular basis of anthocyanin biosynthesis in several organs of the rice plants [50]. In Japan, improved pigmented rice

variety was developed by crossing black rice variety ‘Okunomurasaki’ with high quality-white rice variety ‘Koshihikari’ [137]. In Kazakhstan, adapted pigmented rice variety was developed by crossing pigmented and non-pigmented rice varieties [138]. In Thailand, a new deep purple rice variety ‘Riceberry’ was developed by crossing between non-glutinous purple rice and an aromatic white jasmine rice variety [139-140]. Brazil has been released two advanced pigmented rice varieties; the red rice ‘Rubi’ and the black rice ‘Onix’ [141]. In China, biofortified purple endosperm rice called ‘Zijingmi’ with high anthocyanins concentration was developed by editing anthocyanin biosynthesis [142]. New foods and beverages from pigmented rice also have been developed by using improved processing technologies. This review provides an update information on the genetics, biochemistry and biophysical analysis of anthocyanin in pigmented rice that will facilitate rice breeding program to develop improved pigmented rice varieties.

## 2. Discussion

### 2.1 Anthocyanins in Rice Grains

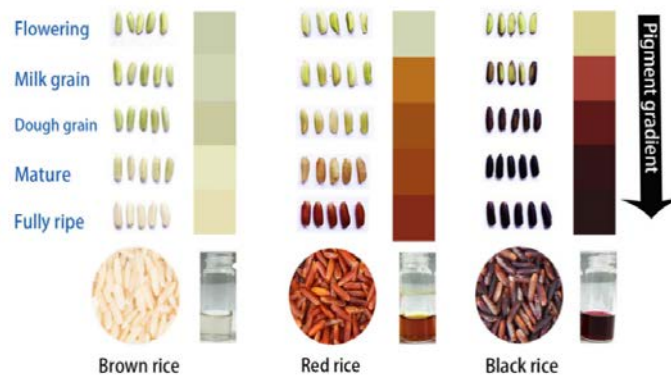
Anthocyanins is mainly accumulated in the pericarp of the rice grains (Figure 1). Purple bran color showed the highest total anthocyanin (2874 cyanidin 3-O-glucoside equivalent (CGE)/100g) followed by black (1884 CGE/100 g), red (8.78 CGE/100g), and brown (3.09 CGE/100g). These rice bran color has been reported to be correlated to the seed dormancy, red bran color rice have a longer dormant compare to the white rice [143]. Based on the quantitative trait loci (QTL) analysis, one QTL qSD7/qPC7 that regulated rice bran color and seed dormancy was identified on chromosome 7 [144]. Anthocyanin contents in the rice pericarp were significantly influenced by environmental conditions and rice developmental stages (Figure 2). Rice developmental stages influence the anthocyanin concentration in caryopsis that shows gradual color changes at each developmental stage. The anthocyanin level increases as the increasing developmental stages and gradual grain filling. At 8-14 days after flowering (DAF), anthocyanins start accumulate in the caryopsis. At the milk stage, caryopsis becomes black and at the maturity stage (35-45 DAF) is the highest anthocyanins concentration accumulate in the caryopsis. The gradual changes of the anthocyanin concentration correlated with the gene expression that control anthocyanin biosynthesis [145]. During maturity stage, the gene expression level of OsDFR, OsF3H, OsAns, and OsCHS are increasing [146]. The anthocyanin accumulation in black rice also influenced by photosynthetic activity.



**Figure 1.** Cross-section of black rice grain showing anthocyanin accumulation in pericarp [66]

Anthocyanin is synthesized on the endoplasmic reticulum, transported through the Golgi apparatus, and accumulated in the vacuole cells of vegetative and generative organs. Anthocyanin biosynthesis is influenced by environmental conditions, such as salinity, drought, abscisic acid (ABA), and rice diseases [147-149]. Black rice pigmentation is regulated by key activator loci for anthocyanin (KALA), such as Kala1, Kala3 or MYB3, and Kala4 [137]. Based on the QTL analysis, Kala1 was identified on chromosome 1 between SSR markers RM7405 and RM7419, Kala3 on chromosome 3 between RM15191 and RM3400, and Kala4 on chromosome 4 between RM1354 and RM7210. LOC\_Os04g0557500 within Kala4 region controls the purple color in the rice pericarp. Anthocyanin concentration in the rice pericarp can be enhanced by overexpress LOC\_ Os04g0557500 [20]. Red rice pericarp is controlled by a QTL rg7.1 on chromosome 7 and LOC\_Os07g11020 was identified within the QTL region [150]. LOC\_Os07g11020 encodes a bHLH TF. Two genes PURPLE PERICARP A (Pp,

Prpa and Prp1) on chromosome 1 and PURPLE PERICARP B (Pb, Prpb and Prp2) on chromosome 4 are regulates the purple color of rice pericarp [151-155].



**Figure 2.** Pigment gradient in brown rice, red rice, and black rice during developmental stages [66]

Transgenic pigmented rice varieties were developed by using transgene stacking system that have higher nutritional and medical values for food and pharmaceutical industries [137]. Anthocyanin concentration in the rice pericarp can be improved by using this genetic engineering technique that can be enhanced their antioxidant activity and seed dormancy period [162]. By enhancing the anthocyanin concentration in the rice pericarp may enhance the abiotic and biotic resistance [50]. Important genes regulating the anthocyanin biosynthesis, such as CHS (chalcone synthase), F3H (flavanone 3]-hydroxylase), DFR (dihydroflavanol), and ANS1 (anthocyanin synthase) were identified by using whole genome sequencing and transcriptomic sequencing in the pigmented rice plants [156]. Pigmented rice produce lower yield and lower grain weight than white rice varieties [157-158]. Lower grain yield of pigmented rice due to the anthocyanin deposition that reduce chlorophyll content in spikelet, decrease photosynthetic rate and also grain filling rate [159]. The accumulation of anthocyanin in pericarp of the pigmented rice cause lower grain weight [145]. The lower grain yield and decreased grain size in black rice near isogenic lines (NILs) population was identified in Japan [137].

## 2.2 Anthocyanins in Rice Floral Organs

Floral organs of the rice plants including stigma and apiculus showing red, purple, or brown color because of anthocyanins accumulation (Figure 3). These obvious color is important in pollination to attract insects and other animals but does not apply in the rice plants due to self-pollination. Anthocyanins content in the floral organs also important as protective agents from ultraviolet radiation and strong light intensity, and also become defense system from abiotic and biotic stresses including salinity, drought, cold, heat, and diseases [160-162]. The specific color of stigma and apiculus is important for rice taxonomy [73], [163-164]. Investigation about purple stigma and apiculus started in 1957 [165]. OsB2 is an important gene regulating anthocyanin biosynthesis in floral organs of the rice plants. The variation color of apiculus is regulated by locus C, OsC1 [50].



**Figure 3.** Purple and white stigma of the rice plants [73]

The variation color of apiculus and stigma are regulated by several QTL regions. Diverse color of apiculus is controlled by C-gene located between SSR markers RM19552-RM19565 [166-167]. Purple apiculus is regulated by Pa-6 and red apiculus is coordinated by OSC [163], [166]. Purple stigma is controlled by Ps-4(t) located in RM253, RM111, and RM6917 [168]. The first purple stigma gene OsC1 was identified on chromosome 6. Based on the map-based cloning strategy, two genes OsC1 and OsDFR that responsible for the purple color of stigma and apiculi were identified in pigmented indica rice cultivar Xieqingzao. A R2R3-MYB transcription factor is encoded by OsC1 on chromosome 6, while a dihydroflavonol 4-reductase is encoded by OsDFR. OsPa gene responsible for apiculi color and OsPs gene regulating the stigma color were identified by transcriptional expression analysis and CRISPR/Cas9. Variety color of stigma and apiculus can be produced by gene interaction of OsC1, OsDFR, OsPa, and OsPs. The purple color of stigma and apiculi is the result of genes interaction OsC1, OsPa or OsPs, and OsDFR. Brown apiculi can be produced by gene interaction OsC1 and OsPa. Knock-out of OsDFR resulting straw-white color stigma [169].

### *2.3. Anthocyanins in Rice Leaves*

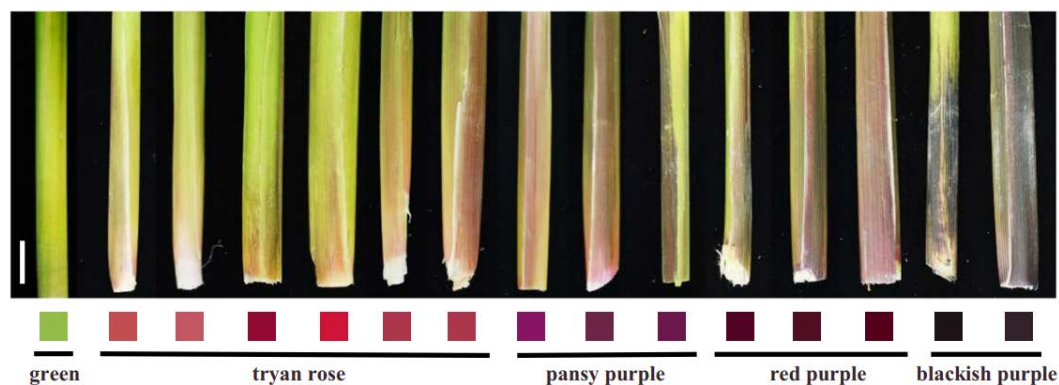
Leaves as the primary organ in photosynthesis promote the biosynthesis of anthocyanin and starch. Anthocyanin accumulation in the rice leaves reduces the efficiency of photosynthetic activity and consequently decrease the rice yield [170]. On the other hand, reducing the anthocyanin concentration in the leaves will increase the photosynthetic activity and subsequently improve rice yield. Consequently, in the rice variety selection process, purple leaf trait become a negative marker [50]. Based on the genomic sequence analysis, accumulation of anthocyanin in the rice leaves is regulated by OsC1 and OsDFR [170]. OsC1 controls cyanidin 3-O-glucoside concentration in the rice leaves [171]. Rb gene on chromosome 1 involves in anthocyanins biosynthesis in the rice leaves was identified by GWAS analysis. LOC\_Os04g0577800 and LOC\_Os04g0616400 on chromosome 4 also involve in anthocyanin biosynthesis in the purple rice leaves that identified by using bulk segregant and transcriptome analysis [172].

### *2.4. Anthocyanins in Rice Leaf Sheath*

Purple leaf sheath in rice due to the accumulation of anthocyanins. Leaf sheath color trait also become a marker in rice variety selection. The level of anthocyanin accumulation in the leaf sheath also influenced by the developmental growth stages. At V4 stage with 4-leaf, the anthocyanin concentration in the leaf sheath ranging from 0.01  $\mu\text{mol/g}$  until 0.06  $\mu\text{mol/g}$ . The highest concentration of anthocyanins starting at active tillering stage until maturity stage around 1.16  $\mu\text{mol/g}$  [73]. Accumulation of the anthocyanins in the leaf sheath also correlated with defense system to the abiotic stresses, including soil acidity, ultraviolet radiation, and temperature [173-174]. Diverse leaf sheath color is regulated by OsC1 gene that has co-segregation with apiculus color [167]. A mutant rice plant Z418 showing purple leaf sheath which was developed from C418 rice variety with green leaf sheet color by modifying OsC1 gene [175]. OsC1 gene also identified in the F2 population of crossing between purple leaf sheath rice (Tainung 72 / TNG72) and green leaf sheath rice (Taichung Sen 17 / TCS17) [176]. There a segregation in that F2 population with ratio 3:1, 3 for purple leaf sheath and 1 for green leaf sheath, indicating that OsC1 is the dominant gene. Based on the RT-PCR analysis, the gene expression of OsC1 in the leaf sheath tissue started at 5-leaf stages [177]. Another gene that regulating the rice purple leaf sheath is PSH1(t) on chromosome 1 that identified in a recombinant inbred line (RIL) population resulting from crossing rice variety IRBB60 and 9407 [152]. Variation of the leaf sheath color also controlled by two QTLs on chromosome 1 and 6 [73], [178-179]. LOC\_Os06g10350 on chromosome 6 as a gene controlling leaf sheath color was also identified by using F2 rice population and 117 markers. LOC\_Os06g10350 belong to the MYB family transcription factor.

Variation of leaf sheath color ranging from light to dark purple showing tyran rose, pansy purple, red purple, and blackish purple. Anthocyanin accumulation in the leaf sheath ranged from 1.04 to 42.77

$\mu\text{mol/g}$ , tyran rose color has the least anthocyanin concentration and blackish purple color has the most anthocyanin content (Figure 4). The diverse leaf sheath color also associated with the rice varieties [73].



**Figure 4.** Diverse rice leaf sheath color [73]

### 2.5. Anthocyanins in Rice Hulls

Colored rice hulls which are black and red due to the accumulation of anthocyanins [36]. About 15% of the rice varieties are colored hull and the most rice varieties (85%) have white-hulled. Colored rice hull responsible to protect rice grain from oxidative stress [180]. China and Japan have been cultivated rice variety with colored hulls since ancient time due to the health positive effects. Recently, rice variety with colored hulls are cultivated in South Asia countries, United States, Italy, and Greece [181]. The type of anthocyanin in the purple rice hull is cyanidin 3-O-glucoside with concentration 2.8 mg/g [182]. Lemma and palea of the rice hull are associated with the rice floral organs and seed characteristics including grain length, grain width, and grain weight [183-188]. The highest anthocyanins accumulation is in purple hulls. Rice hull has been treated as one of residue material from the rice plants, but right now colored rice hull is become antioxidant and anti-cancer sources. Straw-white hull correlated with non-shattered rice grains and became an important marker during rice domestication [189].

Pheno- type	Hull color	Straw-white				Brown		Purple	
						Apiculus	Hull	Apiculus	Hull
Geno- type	C1								
	S1								
	A1								

**Figure 5.** C-S-A gene system of rice hull pigmentation [182]

Methanol extracts of colored rice hulls from rice variety Heuginju with black hull, WD-3 with purple hull, Jeoginju with red hull, and Ilpum with light-brown showed significantly high anti-cancer and antioxidant activities. Black hull of Heuginju showed the highest anti-cancer and antioxidant activities compared to the others colored hulls [190]. Acetone extract of rice hull contain procyanidins [36]. Based on the molecular, genetic, metabolic, and phylogenetic analysis; colored rice hulls were regulated by C-S-A gene system, which C1 encoding MYB transcription factor and become gene that produce color, S1 encoding bHLH protein and acting as a tissue specific regulator, A1 encoding dihydroflavonol reductase and only express when C1 co-ordinate with S1 (Figure 5). Brown hull color is formed when A1 is not expressed. One QTL responsible for black hull trait is on chromosome 4 [191]. Gene Phr1 on chromosome 4 encoding polyphenol oxidase is found to be responsible for black hull color of rice [192-193]. Black hull is also regulated by Bh4 gene on chromosome 4. Another genes that

responsible for black hull are Bh-a, Bh-b, and Bh-c as complementary genes [194]. Two QTLs qHC4 and qHC7 also responsible for black hull coloration, these QTLs were identified by using an F2 population of crossing between SS18-2 and EM93-1 [194].

### 3. Conclusions

Pigmented rice has been popular among rice consumers and increasing demand of pigmented rice have become motivation for rice breeder to develop high yield pigmented rice. Anthocyanin is a source of functional food ingredients, natural colorants, pharmaceuticals, and other industrial biochemical products with the high health benefits. Anthocyanin accumulation in the rice plants can be enhanced by genetic molecular techniques and environmental regulation. Advanced pigmented rice varieties with enhanced anthocyanins content have been developed. To fulfill the consumer increasing demand, it is important to explore deeply the genetic bases of pigmented rice in order to enhance pigmented rice quality, sensory properties, and nutritional content. In this review provides important information for rice breeder to develop high quality pigmented rice based on consumer demand.

### References

- [1] G. B. Gregorio, D. Senadhira, T. Htut, and R. D. Graham, "Improving iron and zinc value of rice for human nutrients," *Agr. Develop.*, 1999, vol. 23, p. 68–87.
- [2] T. Tsuda, M. Watanabe, K. Ohshima, S. Norinobu, S. W. Choi, S. Kawakishi, and T. Osawa, "Antioxidative activity of the anthocyanin pigments cyanidin-3-O- $\beta$ -D-glucoside and cyanidin," *J. Agr. Food Chem.*, 1994, vol. 42, p. 2407–2410.
- [3] Y. M. Chung, J. C. Lee, K. S. Kim, and J. B. Eun, "Chemical compositions of 26 varieties of Korean rice straw," *Food Sci. Biotechnol.*, 2001, vol. 10, pp. 267–271.
- [4] Y. R. Lee, J. Y. Kim, K. S. Woo, I. G. Hwang, K. H. Kim, K. J. Kim, J. H. Kim, and H. S. Jeong, "Changes in the chemical and functional components of Korean rough rice before and after germination," *Food Sci. Biotechnol.*, 2007, vol. 16, p. 1006–1010.
- [5] Y. Dwiningsih, "Molecular genetic analysis of drought resistance and productivity traits of rice genotypes," Ph.D. dissertation, Dept. Crop, Soil, and Env. Sciences, University of Arkansas, Fayetteville, AR, USA, 2020.
- [6] V. C. Ito, A. A. F. Zielinski, I. M. Demiate, M. H. F. Spoto, A. Nogueira, and L. G. Lacerda, "Effects of gamma radiation on the stability and degradation kinetics of phenolic compounds and antioxidant activity during storage of (*Oryza sativa* L.) black rice flour," *Brazilian Archives of Biology and Technology*, 2019, vol. 62, p. 01–14.
- [7] X. Chen, T. Itani, X. Wu, Y. Chikawa, and K. Irifune, "Physiological factors affecting transcription of genes involved in the flavonoid biosynthetic pathway in different rice varieties," *Plant Signaling & Behavior*, 2013, vol. 8, no. 12, p. e27555, doi: 10.4161/psb.27555.
- [8] H. S. Chung and W. S. Woo, "A quinolone alkaloid with antioxidant activity from the aleurone layer of anthocyanin-pigmented rice," *J. Nat. Prod.*, 2001, vol. 64, p. 1579–1580.
- [9] Y. Dwiningsih, A. Kumar, J. Thomas, C. Ruiz, J. Alkahtani, N. Baisakh, and A. Pereira, "Quantitative trait loci and candidate gene identification for chlorophyll content in RIL rice population under drought conditions," *Indonesian Journal of Natural Pigments*, 2021, vol. 3, no. 2, p. 54–64, doi: 10.33479/ijnp.2021.03.2.54
- [10] H. J. Lee, S. K. Oh, H. C. Choi, and S. U. Kim, "Identification of anthocyanins from pigmented rice seed," *Agric. Chem. Biotechnol.*, 1988, vol. 41, p. 257–265.
- [11] K. Yoshinaga, "Liquor with pigments of red rice," *Journal of Brewing Society of Japan*, 1986, vol. 81, p. 337–342.
- [12] W. H. Ling, L. L. Wang, and J. Ma, "Supplementation of the black rice outer fraction to rabbits decrease atherosclerotic plaque formation and increase antioxidant status," *J. Nutr.*, 2002, vol. 132, p. 20–26.



- [13] C. Hu, J. Zawistowski, W. Ling, and D. D. Kitts, "Black rice (*Oryza sativa* L. indica) pigmented fraction suppresses both reactive oxygen species and nitric oxide in chemical and biological model systems," *J. Agr. Food Chem.*, 2003, vol. 51, p. 5271–5277.
- [14] Jr. E. Middleton, C. Kandaswami, and T. C. Theoharides, "The effects of plant flavonoids on mammalian cells: Implications for inflammation, heart disease, and cancer," *Pharmacol. Rev.*, 2000, vol. 52, p. 673–751.
- [15] M. Jang, L. Cai, G. O. Udeani, K. V. Slowing, C. F. Thomas, C. W. Beecher, H. H. Fong, N. R. Farnsworth, A. D. Kinghorn, R. C. Mehta, R. C. Moon, and Pezzuto, "Cancer chemopreventive activity of resveratrol, a natural product derived from grapes," *Science*, 1997, vol. 275, p. 218–220.
- [16] S. I. Shim, J. W. Chung, J. M. Lee, K. T. Hwang, J. Sone, B. S. Hong, H. Y. Cho, and W. J. Jun, "Hepatoprotective effects of black rice on superoxide anion radicals in HepG2 Cells," *Food Sci. Biotechnol.*, 2006, vol. 15, p. 993–996.
- [17] R. C. Chaudhary, "Speciality rices of the world: Effect of WTO and IPR on its production trend and marketing," *Journal of Food, Agriculture and Environment*, 2003, vol. 1, no. 2, p. 34–41.
- [18] Y. Dwiningsih, M. Rahmaningsih, and J. Alkahtani, "Development of single nucleotide polymorphism (SNP) markers in tropical crops," *Adv. Sustain. Sci. Eng. Technol.*, 2020, vol. 2, no. 2, p. 14065.
- [19] S. N. Ryu, S. Z. Park, and C. T. Ho, "High Performance Liquid Chromatographic Determination of Anthocyanin Pigments in Some Varieties of Black Rice," 1998, vol. 6, no. 4, doi: [10.38212/2224-6614.2893](https://doi.org/10.38212/2224-6614.2893).
- [20] T. Oikawa, H. Maeda, T. Oguchi, T. Yamaguchi, N. Tanabe, K. Ebana, M. Yano, T. Ebitani, and T. Izawa, "The Birth of a Black Rice Gene and Its Local Spread by Introgression," *Plant Cell*, 2015, vol. 27, p. 2401–2414.
- [21] P. H. Kristantini, "Potensi pengembangan beras merah sebagai plasma nutfah Yogyakarta," *Jurnal Litbang Pertanian*, 2009, vol. 28, no. 3, p. 88–95.
- [22] R. Pratiwi and Y. A. Purwestri, "Black rice as a functional food in Indonesia," *Functional Foods in Health and Disease*, 2017, vol. 7, no. 3, p. 182–194.
- [23] W. H. Ling, Q. X. Cheng, J. Ma, and T. Wang, "Red and black rice decrease atherosclerotic plaque formation and increase antioxidant status in rabbits," *J. Nutr.*, 2001, vol. 131, p. 1421–1426.
- [24] M. Xia, W. H. Ling, J. Ma, D. D. Kitts, and J. Zawistowski, "Supplementation of diets with the black rice pigment fraction attenuates atherosclerotic plaque formation in apolipoprotein e deficient mice," *The Journal of nutrition*, 2003, vol. 133, no. 3, p. 744–751.
- [25] Z. Xu, N. Hua, and J. S. Godber, "Antioxidant activity of tocopherols, tocotrienols, and  $\gamma$ -Oryzanol components from rice bran against cholesterol oxidation accelerated by 2, 2'-Azobis (2-methylpropionamide) dihydrochloride," *Journal of Agricultural and Food Chemistry*, 2001, vol. 49, no. 4, p. 2077–2081.
- [26] R. Sitrarasi, U. M. Nallal, M. Razia, W. J. Chung, J. Shim, M. Chandrasekaran, Y. Dwiningsih, R. A. Rasheed, J. Alkahtani, M. S. Elshikh, O. Debnath, and Ravindran, "Inhibition of multi-drug resistant microbial pathogens using an ecofriendly root extract of *Furcraea foetida* silver nanoparticles," *Journal of King Saud University-Science*, 2022, vol. 34, no. 2, p. 101794, doi: [10.1016/j.jksus.2021.101794](https://doi.org/10.1016/j.jksus.2021.101794).
- [27] Y. Dwiningsih, "Nutritional characterization of bread contained natural pigment from insect," *Indonesian Scientific Research Olympic*, 2014.
- [28] M. Xia, W. H. Ling, J. Ma, and J. Zawistowski, "Supplementation of diets with the black rice pigment fraction attenuates atherosclerotic plaque formation in apolipoprotein E deficient mice," *J. Nutr.*, 2002, vol. 133, p. 744–751.
- [29] A. N. Chiang, H. L. Wu, H. I. Yeh, C. S. Chu, H. C. Lin, and W. C. Lee, "Antioxidant effects of black rice extract through the induction of superoxide dismutase and catalase activities," *Lipids*, 2006, vol. 41, p. 797–803.
- [30] Y. M. Choi, H. S. Jeong, and J. S. Lee, "Antioxidant activity of methanolic extracts from some grains consumed in Korea," *Food Chem.*, 2007, vol. 103, p. 130–138.

- [31] D. Samyot, A. B. Das, and S. C. Deka, "Pigmented rice a potential source of bioactive compounds: a review," *International Journal of Food Science and Technology*, 2017, vol. 52, p. 1073–1081, doi: 10.1111/ijfs.13378.
- [32] M. Kang, C. W. Rico, and S. Lee, "Varietal Difference in Physicochemical Properties of Pigmented Rice Varieties," *J. Crop Sci. Biotech.*, 2011, doi: 10.1007/s12892-010-0041-8.
- [33] R. Yawadio, S. Tanimori, and N. Morita, "Identification of phenolic compounds isolated from pigmented rices and their aldose reductase inhibitory activities," *Food Chem.*, 2007, vol. 101, p. 1644–1653.
- [34] S. H. Nam, S. P. Choi, M. Y. Kang, H. J. Koh, N. Kozukue, and M. Friedman, "Antioxidative activities of bran extracts from twenty one pigmented rice cultivars," *Food Chem.*, 2006, vol. 94, p. 613–620.
- [35] S. Toyokuni, T. Itani, Y. Morimitsu, K. Okada, M. Ozeki, and S. Kondo, "Protective effect of colored rice over white rice on Fenton reaction-based renal lipid peroxidation in rats," *Free Radic. Res.*, 2002, vol. 36, p. 583–592.
- [36] T. Oki, M. Matsuda, M. Kobayashi, Y. Nishiba, S. Furuta, and I. Suda, "Polymeric procyanidins as radical-scavenging components in red-hulled rice," *J. Agric. Food Chem.*, 2002, vol. 50, p. 7524–7529.
- [37] K. K. Adom and R. H. Liu, "Antioxidant activity of grains," *J. Agric. Food Chem.*, 2002, vol. 50, p. 6170–6182.
- [38] M. C. Lazze, R. Pizzala, M. Savio, L. A. Stivala, E. Prospero, and L. Bianchi, "Anthocyanins protect against DNA damage induced by tert-butyl-hydroperoxide in rat smooth muscle and hepatoma cells," *Genetic Toxicology and Environmental Mutagenesis*, 2003, vol. 535, p. 103–115.
- [39] Y. Dwiningsih, "Margarine with high antioxidant content," *Natural Pigment Conference*, 2012.
- [40] Y. Dwiningsih and S. Notosoedarmo, "Antioxidant sources from insects," *Natural Pigment Conference*, 2012.
- [41] Z. Hou, P. Qin, and G. Ren, "Effect of Anthocyanin-Rich Extract from Black Rice (*Oryza sativa* L. Japonica) on Chronically Alcohol-Induced Liver Damage in Rats," *J. Agric. Food Chem.*, 2010, vol. 58, p. 3191–3196, doi: 10.1021/jf904407x.
- [42] S. Hemamalini, D. S. Umamaheswari, D. R. Lavanya, and R. D. C. Umamaheswara, "Exploring the therapeutic potential and nutritional properties of 'KaruppuKavuni' variety rice of Tamil Nadu," *Int. J. Pharma Bio Sci.*, 2018, vol. 9, p. 88–96.
- [43] I. Konczak and W. Zhang, "Anthocyanins—more than nature's colours," *J. Biomed. Biotechnol.*, 2004, 239–240.
- [44] S. Yazhen, W. Wenju, Z. Panpan, Y. Yuanyuan, D. Panpan, Z. Wusen, and W. Yanling, "Anthocyanins: Novel Antioxidants in Diseases Prevention and Human Health. In *Flavonoids-A Coloring Model for Cheering Up Life*," IntechOpen: London, UK, 2019.
- [45] B. Kocic, S. Filipovic, M. Nikolic, and B. Petrovic, "Effects of anthocyanins and anthocyanin-rich extracts on the risk for cancers of the gastrointestinal tract," *Off. J. Balk. Union Oncol.*, 2011, vol. 16, p. 602–608.
- [46] D. S. Woo, Y. K. Jun, J. P. Mi, H. Sang-Ik, W. K. Hang, R. Ji-Eun, K. O. Sea, H. L. Jin, C. J. Ki, and C. S. You, "Relationship of Radical Scavenging Activities and Anthocyanin Contents in the 12 Colored Rice Varieties in Korea," *J. Korean Soc. Appl. Bi.*, 2011, vol. 54, p. 693–699.
- [47] P. N. Chen, S. C. Chu, H. L. Chiou, C. L. Chiang, S. F. Yang, and Y. Hsieh, "Cancer, Cyanidin 3-glucoside and peonidin 3-glucoside inhibit tumor cell growth and induce apoptosis in vitro and suppress tumor growth in vivo," *J. Nutr.*, 2005, vol. 53, p. 232–243.
- [48] L. P. Luo, B. Han, X. P. Yu, X. Y. Chen, J. Zhou, W. Chen, Y. F. Zhu, X. L. Peng, Q. Zou, and S. Y. Li, "Anti-metastasis activity of black rice anthocyanins against breast cancer: Analyses using an ErbB2 positive breast cancer cell line and tumoral xenograft model," *Asian Pac. J. Cancer Prev.*, 2014, vol. 15, p. 6219–6225.
- [49] S. U. Lule and W. J. F. R. I. Xia, "Food phenolics, pros and cons: A review," *J. Food Rev. Int.*, 2005, vol. 21, p. 367–388.

- [50] D. Xia, H. Zhou, P. Li, B. Wu, Q. Zhang, and Y. He, "How rice organ painted: the genetic basis of anthocyanin biosynthesis in rice," Research Square, 2020, doi: 10.21203/rs.3.rs-101298/v1.
- [51] H. K. Dooner, T. P. Robbins, and R. A. Jorgensen, "Genetics and development control of anthocyanin biosynthesis," Annu. Rev. Genet., 1991, vol. 25, p. 173–199.
- [52] A. Kootstra, "Protection from UV-B-induced DNA damage by flavonoids," Plant Mol. Biol., 1994, vol. 26, p. 771–774.
- [53] V. S. Reddy, K. V. Goud, R. Sharma, and A. R. Reddy, "Ultraviolet-B-responsive anthocyanin production in a rice cultivar is associated with a specific phase of phenylalanine ammonia lyase biosynthesis," Plant Physiol., 1994, vol. 105, p. 1059–1066.
- [54] T. A. Holton and E. C. Cornish, "Genetics and Biochemistry of Anthocyanin Biosynthesis," The Plant Cell, 1995, vol. 7, p. 1071–1083.
- [55] R. A. Dixon and N. Paiva, "Stress-induced phenylpropanoid metabolism," Plant Cell, 1995, vol. 7, p. 1085–1097.
- [56] B. W. Shirley, "Flavonoid biosynthesis: new functions for an old pathway," Trends Plant Sci. 1996, vol. 1, p. 377–382.
- [57] L. Chalker-Scott, "Environmental significance of anthocyanins in plant stress responses," Photochem. Photobiol., 1999, vol. 70, p. 1–9.
- [58] W. B. Shirley, "Biosynthesis of flavonoids and effects of stress," Curr. Opin. Plant Biol., 2002, vol. 5, p. 218–223.
- [59] Y. Dwiningsih and S. Notosoedarmo, "Chlorophyll and Carotenoid in C4, C3, and CAM plants," Indonesian Scientific Research Olympic, 2008.
- [60] C. K. Reddy, L. Kimi, S. Haripriya, and N. Kang, "Effects of polishing on proximate composition, physico-chemical characteristics, mineral composition and antioxidant properties of pigmented rice," Rice Sci., 2017, vol. 24, p. 241–252.
- [61] H. Zhang, Y. Shao, J. Bao, and T. Beta, "Phenolic compounds and antioxidant properties of breeding lines between the white and black rice," Food Chem., 2015, vol. 172, p. 630–639.
- [62] A. Maqsood, Z. I. Khan, K. Ahmad, S. Akhtar, A. Ashfaq, I. S. Malik, R. Sultana, M. Nadeem, J. Alkahtani, Y. Dwiningsih, and M. Elshikh, "Quantitative evaluation of zinc metal in meadows and ruminants for health assessment: implications for humans," Environmental Science and Pollution Research, 2022, vol. 29, p. 21634–21641, doi: 10.1007/s11356-021-17264-1.
- [63] X. Ge, Z. I. Khan, F. Chen, M. Akhtar, K. Ahmad, A. Ejaz, M. A. Ashraf, M. Nadeem, S. Akhtar, J. Alkahtani, Y. Dwiningsih, and M. S. Elshikh, "A study on the contamination assessment, health risk and mobility of two heavy metals in the soil-plants-ruminants system of a typical agricultural region in the semi-arid environment," Environmental Science and Pollution Research, 2021, vol. 29, p. 14584–14594, doi: 10.1007/s11356-021-16756-4.
- [64] Y. Dwiningsih, A. Kumar, J. Thomas, C. Ruiz, J. Alkahtani, A. Al-Hashimi, and A. Pereira, "Identification of Genomic Regions Controlling Chalkiness and Grain Characteristics in a Recombinant Inbred Line Rice Population Based on High-Throughput SNP Markers," Genes, 2021, vol. 12, p. 1690, doi: 10.3390/genes12111690.
- [65] M. N. Clifford, "Anthocyanins: Nature, occurrence and dietary burden," J. Sci. Food Agric., 2000, vol. 80, p. 1063–1072.
- [66] E. Mackon, G. C. Mackon, Y. Ma, M. H. Kashif, N. Ali, B. Usman, and P. Liu, "Recent Insights into Anthocyanin Pigmentation, Synthesis, Trafficking, and Regulatory Mechanisms in Rice (*Oryza sativa* L.) Caryopsis," Biomolecules, 2021, vol 11, p. 394, doi: 10.3390/biom11030394.
- [67] T. Xu, J. Sun, H. Chang, H. Zheng, J. Wang, H. Liu, L. Yang, H. Zhao, and D. T. Zou, "QTL mapping for anthocyanin and proanthocyanidin content in red rice," Euphytica, 2017, vol. 213, p. 243.
- [68] Y. Dwiningsih and S. Notosoedarmo, "Rice contained carotenoid," Indonesian Scientific Research Olympic, 2012.
- [69] M. T. Escribano-Bailón, C. Santos-Buelga, and J. C. Rivas-Gonzalo, "Anthocyanins in cereals," Journal of Chromatography, 2004, vol: A 054, p. 129e141.

- [70] E. S. M. Abdel-Aal, J. C. Young, and I. Rabalski, "Anthocyanin composition in black, blue, pink, purple, and red cereal grains," *Journal of Agricultural and Food Chemistry*, 2006, vol. 54, p. 4696–4704.
- [71] V. S. Reddy, S. Dash, and A. R. Reddy, "Anthocyanin pathway in rice (*Oryza sativa* L.): identification of a mutant showing dominant inhibition of anthocyanins in leaf and accumulation of proanthocyanidins in pericarp," *Theor Appl Genet.*, 1995, vol. 91, p. 301–312.
- [72] W. Sakamoto, T. Ohmori, K. Kageyama, C. Miyazaki, A. Saito, M. Murata, K. Noda, and M. Maekawa, "The purple leaf (Pl) locus of rice: the Plw allele has a complex organization and includes two genes encoding basic helix-loop-helix proteins involved in anthocyanin biosynthesis," *Plant Cell Physiol.*, 2001, vol. 42, p. 982–991.
- [73] H. Chin, Y. Wu, A. Hour, C. Hong, and Y. Lin, "Genetic and evolutionary analysis of purple leaf sheath in rice," *Rice*, 2016, vol. 9, no. 1, p. 8, doi: 10.1186/s12284-016-0080-y.
- [74] Y. Dwiningsih, A. Kumar, J. Thomas, C. Gupta, C. Ruiz, J. Alkahtani, N. Baisakh, and A. Pereira, "Identification and expression of abscisic acid-regulated genes in US RIL rice population under drought conditions," 82<sup>nd</sup> Meeting of Southern Section of the American Society of Plant Biologists, 2021.
- [75] F. D. Goffman and C. J. Bergman, "Rice kernel phenolic content and its relationship with antiradical efficiency," *J. Sci. Food Agric.*, 2004, vol. 84, p. 1235–1240, doi: 10.1002/jsfa.1780.
- [76] X. Q. Chen, N. Nagao, T. Itani, and Irifune, "Anti-oxidative analysis, and identification and quantification of anthocyanin pigments in different coloured rice," *Food Chemistry*, 2012, vol. 135, p. 2783–2788, doi: 10.1016/j.foodchem.2012.06.098.
- [77] R. Koes, W. Verweij, and F. Quattrocchio, "Flavonoids: a colorful model for the regulation and evolution of biochemical pathways," *Trends Plant Sci.*, 2005, vol. 10, p. 1360–1385.
- [78] W. S. Brenda, "Flavonoid biosynthesis. A colorful model for genetics, biochemistry, cell biology, and biotechnology," *Plant Physiol.*, 2001, vol. 126, p. 485–493.
- [79] P. Loypimai, A. Moongngarm, and P. Chottanom, "Thermal and pH degradation kinetics of anthocyanins in natural food colorant prepared from black rice bran," *Journal of Food Science and Technology*, 2016, vol. 53, no. 1, p. 461–470.
- [80] Y. Zhang, X. S. Hu, F. Chen, J. H. Wu, X. J. Liao, and Z. F. Wang, "Stability and color characteristics of PET-treated cyanidin-3-glucoside during storage," *Food Chemistry*, 2008, vol. 106, no. 2, p. 669–676.
- [81] B. K. Tiwari, C. P. O' Donnell, K. Muthukumarappan, and P. J. Cullen, "Anthocyanin and color degradation in ozone treated blackberry juice," *Innovative Food Science and Emerging Technologies*, 2009, vol. 10, no. 1, p. 70–75.
- [82] G. Gradinaru, C. G. Biliaderis, S. Kallithraka, P. Kefalas, and C. Garcia-Viguera, "Thermal stability of *Hibiscus sabdariffa* L. anthocyanins in solution and in solid state: Effects of copigmentation and glass transition," *Food Chemistry*, 2003, vol. 83, no. 3, p. 423–436.
- [83] Y. Dwiningsih, A. Kumar, J. Thomas, S. Yingling, and A. Pereira, "Identification of QTLs associated with drought resistance traits at reproductive stage in K/Z RILs rice population," 5<sup>th</sup> Annual Meeting of the Arkansas Bioinformatics Consortium AR-BIC 2020, Bioinformatics in Food and Agriculture, 2020.
- [84] Y. Dwiningsih, "Thermostability of natural pigment from secang bark (*Caesalpinia sappan* Linn.)," Indonesian Scientific Research Olympic, 2013.
- [85] Z. Hou, P. Qing, and G. Ren, "Effect of anthocyanin-rich extract from black rice (*Oryza sativa* L. Japonica) on chronically alcohol-induced liver damage in rats," *Journal of Agricultural and Food Chemistry*, 2010, vol. 58, p. 3191e3196.
- [86] J. Shipp and Abdel-Aal, "Food Applications and Physiological Effects of Anthocyanins as Functional Food Ingredients," *The Open Food Science Journal*, 2010, vol. 4, p. 7–22.
- [87] Q. G. Tian, M. M. Giusti, G. D. Stoner, and S. J. Schwartz, "Screening for anthocyanins using high-performance liquid chromatography coupled to electrospray ionization tandem mass spectrometry

- with precursor-ion analysis, product-ion analysis, common-neutral-loss analysis, and selected reaction monitoring,” *Journal of Chromatography A*, 2005, vol. 1091, p. 72–82.
- [88] Y. Yao, W. Sang, M. J. Zhou, and G. X. Ren, “Antioxidant and  $\alpha$ -glucosidase activity of colored grains in China,” *Journal of Agricultural and Food Chemistry*, 2010, vol. 58, p. 770–774.
- [89] J. H. Lee, “Identifications and quantification of anthocyanins from the grains of black rice (*Oryza sativa* L.) varieties,” *Food Science and Biotechnology*, 2010, vol. 19, p. 391e397.
- [90] M. Hiemori, E. Koh, and A. E. Mitchell, “Influence of cooking on anthocyanins in black rice (*Oryza sativa* L. *japonica* var. SBR),” *J. Agric. Food Chem.*, 2009, vol. 57, p. 1908–1914, doi: 10.1021/jf803153z.
- [91] H. Yoshida, Y. Tomiyama, and Y. Mizushina, “Lipid components, fatty acids and triacylglycerol molecular species of black and red rices,” *Food Chemistry*, 2010, vol. 123, p. 210e215.
- [92] L. Kong, Y. Wang, and Y. Cao, “Determination of Myo-inositol and D-chiroinositol in black rice bran by capillary electrophoresis with electrochemical detection,” *Journal of food composition and analysis*, 2008, vol. 21, no. 6, p. 501–504.
- [93] S. J. Han, S. N. Ryu, and S. S. Kang, “A New 2-Arylbenzofuran with Antioxidant Activity from the Black Colored Rice (*Oryza sativa* L.) Bran,” *Chem. Pharm. Bull.*, 2004, vol. 52, no. 11, p. 1365–1366.
- [94] Y. Dwiningsih and S. Notosoedarmo, “Rice contained carotenoid,” *Indonesian Scientific Research Olympic*, 2012.
- [95] U. K. S. Kushwaha, “Black Rice: Research History and Development,” *Springer International Publishing*, 2016, p. 21–31.
- [96] J. Surh and E. Koh, “Effects of four different cooking methods on anthocyanins, total phenolics and antioxidant activity of black rice,” *Journal of the Science of Food and Agriculture*, 2014, vol. 94, no. 15, p. 3296–3304.
- [97] Y. Dwiningsih and S. Notosoedarmo, “Potential of green microalgae (*Haematococcus pluvialis*) as food supplement,” *Indonesian Scientific Research Olympic*, 2012.
- [98] S. Duyi, A. Baran, and D. S. Chandra, “Pigmented rice a potential source of bioactive compounds: A review,” *International Journal of Food Science & Technology*, 2017, vol. 52, no. 5, p. 1073–1081.
- [99] T. L. Jeng, C. C. Lai, P. T. Ho, Y. J. Shih, and J. M. Sung, “Agronomic, molecular and antioxidative characterization of red- and purple-pericarp rice (*Oryza sativa* L.) mutants in Taiwan,” *Journal of Cereal Science*, 2012, vol. 56, no. 2, p. 425–431.
- [100] Y. Dwiningsih, J. Thomas, A. Kumar, C. Gupta, C. Ruiz, S. Yingling, E. Crowley, and A. Pereira, “Molecular genetic analysis of drought resistance and productivity mechanisms in rice,” *Plant and Animal Genome XXVIII Conference*, January 11-15, 2020.
- [101] M. Kim, H. Kim, K. Koh, H. Kim, Y. S. Lee, and Y. H. Kim, “Identification and quantification of anthocyanin pigments in colored rice,” *Nutrition Research and Practice*, 2008, vol. 2, no. 1, p. 46–49.
- [102] E. G. N. Mbanjo, T. Kretschmar, H. Jones, N. Ereful, C. Blanchard, L. A. Boyd, and N. Sreenivasulu, “The Genetic Basis and Nutritional Benefits of Pigmented Rice Grain,” *Front. Genet.*, 2020, vol. 11, no. 229, doi: 10.3389/fgene.2020.00229.
- [103] W. A. Hurtada, A. S. A. Barrion, and M. F. R. Nguyen-Orca, “Mineral content of dehulled and well-milled pigmented and non-pigmented rice varieties in the Philippines,” *Int. Food Res. J.*, 2018, vol. 25, p. 2063–2067.
- [104] Y. Shao, Z. Hu, Y. Yu, R. Mou, Z. Zhu, and T. Beta, “Phenolic acids, anthocyanins, proanthocyanidins, antioxidant activity, minerals and their correlations in non-pigmented, red, and black rice,” *Food Chemistry*, 2018, vol. 239, p. 733–741.
- [105] R. S. Raghuvanshi, A. Dutta, G. Tewari, and S. Suri, “Qualitative characteristics of red rice and white rice procured from local market of Uttarakhand?: a comparative study,” *J. Rice Res.*, 2017, vol. 10, p. 49–53.

- [106] M. I. Hashmi and J. S. Tianlin, "Minerals contents of some indigenous rice varieties of Sabah Malaysia," *Int. J. Agric. For. Plant.*, 2016, vol. 2, p. 31–34.
- [107] Y. Dwiningsih, A. Kumar, J. Thomas, S. Yingling, and A. Pereira, "Molecular genetic analysis of drought resistance and productivity in K/Z RIL rice population," *Arkansas Bioinformatics Consortium*, 2019.
- [108] T. Frank, B. Reichardt, Q. Shu, and K. Engel, "Metabolite profiling of colored rice (*Oryza sativa* L.) grains," *Journal of Cereal Sciences*, 2012, vol. 55, p. 112–119, doi: 10.1016/j.jcs.2011.09.009.
- [109] Y. Morimitsu, K. Kubota, T. Tashiro, E. Hashizume, T. Kamiya, and T. Osawa, "Inhibitory effect of anthocyanins and colored rice on diabetic cataract formation in the rat lenses," *International Congress Series*, 2002, vol. 1245, p. 503–508.
- [110] M. Frei and K. Becker, "Fatty acids and all-trans- $\beta$ -carotene are correlated in differently colored rice landraces," *J Sci Food Agric.*, 2005, vol. 85, p. 2380–2384, doi: 10.1002/jsfa.2263
- [111] J. Y. Kim, M. H. Do, and S. S. Lee, "The effects of a mixture of brown and black rice on lipid profiles and antioxidant status in rats," *Annals of Nutrition and Metabolism*, 2006, vol. 50, p. 347e353.
- [112] Y. Dwiningsih, "Production of secang bark (*Caesalpinia sappan* Linn.) as natural pigment for food with spray drying method," *Indonesian Scientific Research Olympic*, 2014.
- [113] Z. Hou, P. Qin, Y. Zhang, S. Cui, and G. Ren, "Identification of anthocyanins isolated from black rice (*Oryza sativa* L.) and their degradation kinetics," *Food Research International*, 2013, vol. 50, no. 2, p. 691–697.
- [114] Y. Dwiningsih, J. Thomas, A. Kumar, C. Gupta, E. Crowley, C. Ruiz, and A. Pereira, "Drought stress response in US recombinant inbred line of rice population," *National Science Foundation (NSF) 26<sup>th</sup> National Conference*, 2019, vol. 26, no. 76, p. 127.
- [115] J. Ma, Y. Li, and J. Li, "Constituents of red yeast rice, a traditional Chinese food and medicine," *J Agric Food Chem.*, 2000, vol. 48, p. 5220–5225.
- [116] R. E. Wrolstad, "Anthocyanin pigments – bioactivity and coloring properties," *J. Food Sci.*, 2004, vol. 69, p. C419-C425.
- [117] F. C. Stintzing and R. Carle, "Functional properties of anthocyanins and betalins in plants, food, and in human nutrition," *Trends Food Sci. Technol.*, 2004, vol. 15, p. 19–38.
- [118] M. H. Ali, M. I. Khan, S. Bashir, M. Azam, M. Naveed, R. Qadri, S. Bashir, F. Mehmood, M. A. Shoukat, and Y. Li, "Biochar and *Bacillus* sp. MN54 Assisted Phytoremediation of Diesel and Plant Growth Promotion of Maize in Hydrocarbons Contaminated Soil," *Agronomy*, 2021, vol. 11, p. 1795, doi: 10.3390/agronomy11091795.
- [119] H. E. Khoo, A. Azlan, S. T. Tang, and S. M. Lim, "Anthocyanidins and anthocyanins: Colored pigments as food, pharmaceutical ingredients, and the potential health benefits," *Food Nutr. Res.*, 2017, vol. 61, p. 1361779.
- [120] M. Turturică, A. M. Oancea, G. Râpeanu, and G. Bahrim, "Anthocyanins: Naturally occurring fruit pigments with functional properties," *Ann. Univ. Dunarea Jos Galati Fascicle VI-Food Technol.*, 2015, vol. 39, p. 9–24.
- [121] T. A. Holton and E. C. Cornish, "Genetics and Biochemistry of Anthocyanin Biosynthesis," *The Plant Cell*, 1995, vol. 7, p. 1071–1083.
- [122] M. W. Zhang, R. F. Zhang, F. X. Zhang, and R. H. Liu, "Phenolic Profiles and Antioxidant Activity of Black Rice Bran of Different Commercially Available Varieties," *J. Agric. Food Chem.*, 2010, vol. 58, p. 7580–7587, doi: 10.1021/jf1007665
- [123] Y. Pang, S. Ahmed, Y. Xu, T. Beta, Z. Zhu, Y. Shao, and J. Bao, "Bound phenolic compounds and antioxidant properties of whole grain and bran of white, red and black rice," *Food Chemistry*, 2018, vol. 240, p. 212–221.
- [124] Y. Dwiningsih, "Spectroscopy and natural pigment structural determination," *Natural Pigment Conference*, 2012.

- [125] Y. Dwiningsih, F. S. Rondonuwu, and M. Martosupono, "The role of curcumin in bacteriochlorophyll a activity", Satya Wacana Christian University, Salatiga, Jawa Tengah, Indonesia, 2009.
- [126] A. Rafael, Y. Dwiningsih, H. Tuririday, and F. F. Karwur, "Natural colorant and biopsychology. Biopsychology: Improving the Quality of Life, First National Conference on Biopsychology," Gadjah Mada University, Yogyakarta, Indonesia, 2009.
- [127] F. Finocchiaro, B. Ferrari, and A. Gianinetti, "A study of biodiversity of flavonoid content in the rice caryopsis evidencing simultaneous accumulation of anthocyanins and proanthocyanidins in a black-grained genotype," *Journal of Cereal Science*, 2010, vol. 51, p. 28e34.
- [128] T. Furukawa, M. Maekawa, T. Oki, I. Suda, S. Lida, H. Shimada, I. Takamura, and K. Kadowaki, "The Rc and Rd genes are involved in proanthocyanidin synthesis in rice pericarp," *Plant Journal*, 2006, vol. 49, p. 91e102.
- [129] H. S. Chung and W. S. Woo, "A quinolone alkaloid with antioxidant activity from the aleurone layer of anthocyanin-pigmented rice," *J. Nat. Prod.*, 2001, vol. 64, p. 1579–1580.
- [130] J. W. Hyun and H. S. Chung, "Cyanidin and Malvidin from *Oryza sativa* cv. Heugjinjubyeo Mediate Cytotoxicity against Human Monocytic Leukemia Cells by Arrest of G2/M Phase and Induction of Apoptosis," *J. Agric. Food Chem.*, 2004, vol. 52, p. 2213–2217, doi: 10.1021/jf030370h
- [131] Y. Li, F. Teng, F. Shi, L. Wang, and Z. Chen, "Effects of high-temperature air fluidization (HTAF) on eating quality, digestibility, and antioxidant activity of black rice (*Oryza sativa* L.)," *Starch – Stärke*, 2017, vol. 69, p. 7–8, doi: 1600274-n/a.
- [132] T. Laokuldilok and N. Kanha, "Effects of processing conditions on powder properties of black glutinous rice (*Oryza sativa* L.) bran anthocyanins produced by spray drying and freeze drying," *LWT – Food Science and Technology*, 2015, vol. 64, no. 1, p. 405–411.
- [133] L. Meng, W. Zhang, Z. Wu, A. Hui, H. Gao, P. Chen, and Y. He, "Effect of pressure-soaking treatments on texture and retrogradation properties of black rice," *LWT – Food Science and Technology*, 2018, vol. 93, p. 485–490.
- [134] Y. Dwiningsih, A. Kumar, J. Thomas, S. Yingling, and A. Pereira, "Molecular genetic analysis of drought resistance and productivity in US rice cultivars," *Plant and Animal Genome XXVII Conference (January 12-16, 2019)*.
- [135] J. D. Arbelaez, L. T. Moreno, N. Singh, C. W. Tung, L. G. Maron, and Y. Ospina, "Development and GBS-genotyping of introgression lines (ILs) using two wild species of rice. *O. meridionalis* and *O. rufipogon*, in a common recurrent parent, *O. sativa* cv. Curinga," *Mol. Breed.*, 2015, vol. 35, no. 81, doi: 10.1007/s11032-015-0276-7.
- [136] M. A. R. Bhuiyan, M. K. Narimah, H. A. Rahim, M. Z. Abdullah, and R. Wickneswari, "Transgressive variants for red pericarp grain with high yield potential derived from *Oryza rufipogon* × *Oryza sativa*: field evaluation, screening for blast disease, QTL validation and background marker analysis for agronomic traits," *Field Crops Res.*, 2011, vol. 121, p. 232–239, doi: 10.1016/j.fcr.2010.12.012.
- [137] H. Maeda, T. Yamaguchi, M. Omoteno, T. Takarada, K. Fujita, and K. Murata, "Genetic dissection of black grain rice by the development of a near isogenic line," *Breed. Sci.*, 2014, vol. 64, p. 134–141, doi: 10.1270/jsbbs.64.134.
- [138] A. B. Rysbekova, D. T. Kazkeyev, B. N. Usenbekov, Z. M. Mukhina, E. A. Zhanbyrbaev, and I. Sartbaeva, "Prebreeding selection of rice with colored pericarp based on genotyping Rc and Pb genes," *Russ. J. Genet.*, 2017, vol. 53, p. 49–58, doi: 10.1134/s1022795416110119.
- [139] P. Waiyawuththanapoom, W. Waiyawuththanapoom, and P. Tirastittam, "Social media as a channel for Thailand's Rice Berry Product," *Int. J. Econ. Manag. Eng.*, 2015, vol. 9, p. 904–907.
- [140] Gene Discovery Rice and Rice Science Center. Riceberry. (2017). Accessed: March 15, 2022. [Online]. Available: <http://dna.kps.ku.ac.th/index.php/news-articles-rice-rsc-rgdu-knowledge/rice-breeding-lab/riceberry-variety>

- [141] E. Wickert, M. A. Schiocchet, J. A. Noldin, J. V. Raimondi, A. De Andrade, and K. K. Scheuermann, "Exploring variability?: New Brazilian varieties SCS119 Rubi and SCS120 Onix for the specialty rices market," *Open J. Genet.*, 2014, vol. 4, p. 157–165, doi: 10.4236/ojgen.2014.42016
- [142] Q. Zhu, S. Yu, D. Zeng, H. Liu, H. Wang, Z. Yang, X. Xie, R. Shen, J. Tan, H. Li, X. Zhao, Q. Zhang, Y. Chen, J. Guo, L. Chen, and Y. G. Liu, "Development of "Purple Endosperm Rice" by Engineering Anthocyanin Biosynthesis in the Endosperm with a High-Efficiency Transgene Stacking System," *Mol. Plant*, 2017, vol. 10, p. 918–929.
- [143] X. Y. Gu, S. F. Kianian, G. A. Hareland, B. L. Hoffer, and M. E. Foley, "Genetic analysis of adaptive syndromes interrelated with seed dormancy in weedy rice (*Oryza sativa*)," *Thero. Appl. Genet.*, 2005, vol. 110, p. 1108–1118.
- [144] X. Y. Gu, M. E. Foley, D. P. Horvath, J. V. Anderson, J. Feng, L. Zhang, C. R. Mowry, H. Ye, J. C. Suttle, K. Kadowaki, and Z. Chen, "Association between seed dormancy and pericarp color is controlled by a pleiotropic gene that regulates abscisic acid and flavonoid synthesis in weedy red rice," *Genetics*, 2011, vol. 189, p. 1515–1524.
- [145] M. M. Rahman, K. E. Lee, and S. G. Kang, "Allelic Gene Interaction and Anthocyanin Biosynthesis of Purple Pericarp Trait for Yield Improvement in Black Rice," *Journal of Life Science*, 2016, vol. 26, p. 727–736, doi: 10.5352/JLS.2016.26.6.727.
- [146] B. G. Kim, J. H. Kim, S. Y. Min, K. Shin, J. H. Kim, H. Y. Kim, S. N. Ryu, and J. Ahn, "Anthocyanin content in rice is related to expression levels of anthocyanin biosynthetic genes," *Plant Biol.*, 2007, vol. 50, p. 156–160.
- [147] N. Ithal and A. R. Reddy, "Rice flavonoid pathway genes, OsDfr and OsAns, are induced by dehydration, high salt and ABA, and contain stress responsive promoter elements that interact with the transcription activator, OsC1-MYB," *Plant Sci.*, 2004, vol. 166, p. 1505–1513.
- [148] M. Gandikota, D. A. Kochko, L. L. Chen, N. Ithal, C. Fauquet, and A. R. Reddy, "Development of transgenic rice plants expressing maize anthocyanin genes and increased blast resistance," *Mol. Breeding*, 2001, vol. 7, p. 73–83.
- [149] Y. Dwiningsih, J. Thomas, A. Kumar, C. Gupta, S. Yingling, S. Basu, and A. Pereira, "Circadian expression patterns of the HYR gene," *Arkansas Bioinformatics Consortium*, 2018, vol. 7, no. 11, p. 34.
- [150] E. M. Septiningsih, K. R. Trijatmiko, S. Moeljopawiro, and S. R. McCouch, "Identification of quantitative trait loci for grain quality in an advanced backcross population derived from the *Oryza sativa* variety IR64 and the wild relative *O. rufipogon*," *Thero. Appl. Genet.*, 2003, vol. 107, p. 1433–1441.
- [151] Y. Shao, L. Jin, G. Zhang, Y. Lu, Y. Shen, and J. Bao, "Association mapping of grain color, phenolic content, flavonoid content and antioxidant capacity in dehulled rice," *Thero. Appl. Genet.*, 2011, vol. 122, p. 1005–1016.
- [152] W. Y. Wang, H. F. Ding, G. X. Li, M. S. Jiang, R. F. Li, X. Liu, Y. Zhang, and F. Y. Yao, "Delimitation of the PSH1(t) gene for rice purple leaf sheath to a 23.5 kb DNA fragment," *Genome*, 2009, vol. 52, p. 268–274.
- [153] C. X. Wang and Q. Y. Shu, "Fine mapping and candidate gene analysis of purple pericarp gene Pb in rice (*Oryza sativa* L.)," *Chin. Sci. Bull.*, 2007, vol. 52, p. 3097–3104.
- [154] J. P. Hu, B. Anderson, and S. R. Wessler, "Isolation and Characterization Of Rice R Genes Evidence for Distinct Evolutionary Paths in Rice and Maize," *Genetics*, 1996, vol. 142, p. 1021–1031.
- [155] A. C. Mathilde, M. F. Theresa, G. C. Yong, N. A. Sang, C. Julapark, K. S. Wu, J. H. Xiao, Z. H. Yu, C. R. Pamela, E. H. Sandra, S. Gerard, R. M. Susan, and D. T. Steven, "Saturated Molecular Map of the Rice Genome Based on an Interspecific Backcross Population," *Genetics*, 1994, vol. 138, p. 1251–1274.



- [156] J. H. Oh, Y. J. Lee, E. J. Byeon, B. C. Kang, D. S. Kyeoung, and C. K. Kim, “Whole-genome resequencing and transcriptomic analysis of genes regulating anthocyanin biosynthesis in black rice plants,” *3 Biotech.*, 2018, vol. 8, p. 115.
- [157] Z. M. Peng, M. W. Zhang, and J. M. Tu, “Breeding and nutrient evaluation on three-lines and their combination of indica black glutinous rice hybrid,” *Acta Agronomica Sinica*, 2004, vol. 30, p. 342
- [158] H. H. Yoon, Y. S. Paik, J. B. Kim, and T. R. Hahn, “Identification of anthocyanidins from Korean pigmented rice,” *Agric. Chem. Biotechnol.*, 1995, vol. 38, p. 581–583.
- [159] M. M. Rahman, K. E. Lee, and S. G. Kang, “Studies on the effects of pericarp pigmentation on grain development and yield of black rice,” *Indian J. Genet.*, 2015, vol. 75, p. 426–433.
- [160] K. Petroni and C. Tonelli, “Recent advances on the regulation of anthocyanin synthesis in reproductive organs,” *Plant Sci.*, 2011, vol. 181, no. 3, p. 219–229, doi: 10.1016/j.plantsci.2011.05.009
- [161] K. Lin-Wang, K. Bolitho, K. Grafton, A. Kortstee, S. Karunairetnam, T. K. McGhie, R. V. Espley, R. P. Hellens, and A. C. Allan, “An R2R3 MYB transcription factor associated with regulation of the anthocyanin biosynthetic pathway in Rosaceae,” *BMC Plant Biol.*, 2010, vol. 10, no. 1, p. 50, doi: 10.1186/1471-2229-10-50
- [162] Y. Dwiningsih, “Lichenes: pigmented pollution bioindicator,” *BioS Popular Biology Magazine*, Satya Wacana Christian University, 2010.
- [163] S. Zhao, C. Wang, J. Ma, S. Wang, P. Tian, J. Wang, Z. Cheng, X. Zhang, X. Guo, and C. Lei, “Map-based cloning and functional analysis of the chromogen gene C in rice (*Oryza sativa* L.),” *J Plant Biol.*, 2016, vol. 59, no. 5, p. 496–505, doi: 10.1007/s12374-016-0227-9
- [164] Y. Dwiningsih and S. Notosoedarmo, “Carotenoid biosynthesis in the anther”, *Natural Pigment Conference*, 2012.
- [165] M. Takashi, “Analysis on apiculus color genes essential to anthocyanin coloration in rice,” *Agr Hokkaido Univ.*, 1957, vol. 50, p. 266–362.
- [166] X. Liu, X. Sun, W. Wang, H. Ding, W. Liu, G. Liu, M. Jiang, C. Zhu, and F. Yao, “Fine mapping of Pa-6 gene for purple apiculus in rice,” *J. Plant. Biol.*, 2012, vol. 55, p. 218–225.
- [167] F. J. Fan, Y. Y. Fan, J. H. Du, and J. Y. Zhuang, “Fine Mapping of C (Chromogen for Anthocyanin) Gene in Rice,” *Rice Science*, 2008, vol. 15, p. 1–6.
- [168] Z. Chen, W. Deng, F. Li, J. Zhou, J. Li, P. Xu, X. Deng, F. Hu, L. Wang, and S. Chen, “A genetic study on the purple stigma genes and their locations in *Oryza longistaminata*,” *J. Yunnan Univ.-Nat. Sci. Ed.*, 2010, vol. 32, p. 103–107.
- [169] L. Meng, C. Qi, C. Wang, S. Wang, C. Zhou, Y. Ren, Z. Cheng, X. Zhang, X. Guo, Z. Zhao, J. wang, Q. Lin, S. Zhu, H. Wang, Z. Wang, C. Lei, and J. Wan, “Determinant Factors and Regulatory Systems for Anthocyanin Biosynthesis in Rice Apiculi and Stigmas,” *Rice*, 2021, vol. 14, no. 37, doi: 10.1186/s12284-021-00480-1
- [170] J. Zheng, H. Wu, H. Zhu, C. Huang, C. Liu, Y. Chang, Z. Kong, Z. Zhou, G. Wang, Y. Lin, and H. Chen, “Determining factors, regulation system, and domestication of anthocyanin biosynthesis in rice leaves,” *New Phytol.*, 2019, vol. 223, no. 2, p. 705–721, doi: 10.1111/nph.15807
- [171] C. Chen, W. Wu, X. Sun, B. Li, G. Hu, Q. Zhang, J. Li, H. Zhang, and Z. Li, “Fine-mapping and candidate gene analysis of BLACK HULL1 in rice (*Oryza sativa* L.),” *J. Plant Omics.*, 2014, vol. 7, no. 12.
- [172] J. Gao, G. Dai, W. Zhou, H. Liang, J. Huang, D. Qing, W. Chen, H. Wu, X. Yang, and D. J. Li, “Mapping and identifying a candidate gene Plr4, a recessive gene regulating purple leaf in rice, by using bulked segregant and transcriptome analysis with next-generation sequencing,” *Int. J. Mol. Sci.*, 2019, vol. 20, p. 4335.
- [173] K. R. Reddy, V. G. Kakani, D. Zhao, S. Koti, and W. Gao, “Interactive effects of ultraviolet-B radiation and temperature on cotton physiology, growth, development and hyperspectral reflectance,” *Photochem Photobiol.*, 2004, vol 79, p. 416–427.

- [174] Y. Dwiningsih, J. Mangimbulude, and A. S. Krave, "Leachate denitrification activity from Semarang Jatibarang landfill," Satya Wacana Christian University, Salatiga, Jawa Tengah, Indonesia, 2007.
- [175] D. Gao, B. He, Y. Zhou, and L. Sun, "Genetic and molecular analysis of a purple sheath somaclonal mutant in japonica rice," *Plant Cell Reports*, 2011, vol. 30, p. 901–911.
- [176] H. Chin, Y. Wu, A. Hour, C. Hong, and Y. Lin, "Genetic and evolutionary analysis of purple leaf sheath in rice," *Rice*, 2016, vol. 9, no. 1, p. 8, doi: 10.1186/s12284-016-0080-y.
- [177] A. Procissi, S. Dolfini, A. Ronchi, and C. Tonelli, "Light-dependent spatial and temporal expression of regulatory genes in developing maize seeds," *Plant Cell*, 1997, vol. 9, p. 1547–1557.
- [178] B. Yue, K. H. Cui, S. B. Yu, W. Y. Xue, L. J. Luo, and Y. Z. Xing, "Molecular marker-assisted dissection of quantitative trait loci for seven morphological traits in rice (*Oryza sativa* L.)," *Euphytica*, 2006, vol. 150, p. 131–139.
- [179] B. N. Hadagal, A. Manjunath, and J. V. Goud, "Linkage of genes for anthocyanin pigmentation in rice (*Oryza sativa* L.)," *Euphytica*, 1980, vol. 30, p. 747–754.
- [180] S. Lee, J. Kim, S. Jeong, D. Kim, J. Ha, K. Nam, and D. Ahn, "Effect of Far-Infrared Radiation on the Antioxidant Activity of Rice Hulls," *J. Agric. Food Chem.*, 2003, vol. 51, p. 4400–4403, doi: 10.1021/jf0300285.
- [181] D. Simmons and R. Williams, "Dietary practices among Europeans and different South Asian groups in Coventry," *Br. J. Nutr.*, 1997, vol. 78, p. 5–14.
- [182] X. Sun, Z. Zhang, C. Chen, W. Wu, N. Ren, C. Jang, J. Yu, Y. Zhao, X. Zheng, Q. Yang, H. Zhang, J. Li, and Z. Li, "The C–S–A gene system regulates hull pigmentation and reveals evolution of anthocyanin biosynthesis pathway in rice," *Journal of Experimental Botany*, 2018, vol. 69, no. 7, p. 1485–1498, doi: 10.1093/jxb/ery001.
- [183] X. Li, L. Sun, L. Tan, F. Liu, Z. Zhu, Y. Fu, X. Sun, X. Sun, D. Xie, and C. Sun, "TH1, a DUF640 domain-like gene controls lemma and palea development in rice," *Plant Mol. Biol.* 2012, vol. 78, p. 351–359, doi: 10.1007/s11103-011-9868-8.
- [184] J. F. Weng, S. H. Gu, and X. Y. Wan, "Isolation and initial characterization of GW5, a major QTL associated with rice grain width and weight," *Cell Res.*, 2008, vol. 18, p. 1199–1209.
- [185] A. Shomura, T. Izawa, K. Ebana, T. Ebitani, H. Kanegae, S. Konishi, and M. Yano, "Deletion in a gene associated with grain size increased yields during rice domestication," *Nat. Genet.*, 2008, vol. 40, p. 1023–1028.
- [186] X. J. Song, W. Huang, M. Shi, M. Z. Zhu, and H. X. Lin, "A QTL for rice grain width and weight encodes a previously unknown RINGtype E3 ubiquitin ligase," *Nat. Genet.*, 2007, vol. 39, p. 623–630.
- [187] C. C. Fan, Y. Z. Xing, H. L. Mao, T. T. Lu, B. Han, C. G. Xu, X. H. Li, and Q. F. Zhang, "GS3, a major QTL for grain length and weight and minor QTL for grain width and thickness in rice, encodes a putative transmembrane protein," *Theor. Appl. Genet.*, 2006, vol. 112, p. 1164–1171.
- [188] Y. Dwiningsih, A. Kumar, J. Thomas, and A. Pereira, "Identification drought-tolerance rice variety for reducing climatic impacts on rice production," Fulbright Enrichment Seminar Climate Change, Estes Park, Colorado, USA, 2017.
- [189] B. Zhu, L. Si, Z. Wang, Y. Zhou, J. Zhu, Y. Shangguan, D. Lu, D. Fan, C. Li, H. Lin, Q. Qian, T. Sang, B. Zhou, Y. Minobe, and B. Han, "Genetic Control of a Transition from Black to Straw-White Seed Hull in Rice Domestication," *Plant Physiology*, 2011, vol. 155, p. 1301–1311, doi: 10.1104/pp.110.168500.
- [190] J. A. Baek, N. J. Chung, K. C. Choi, J. M. Hwang, and J. C. Lee, "Hull extracts from pigmented rice exert antioxidant effects associated with total flavonoid contents and induce apoptosis in human cancer cells," *Food Sci. Biotechnol.*, 2015, vol. 24, p. 241–247.
- [191] W. Chen, Y. Q. Gao, W. B. Xie, L. Gong, K. Lu, W. S. Wang, Y. Li, X. Q. Liu, H. Y. Zhang, H. X. Dong, W. Zhang, L. J. Zhang, S. Yu, G. W. Wang, X. M. Lian, and J. Luo, "Genome-wide association analyses provide genetic and biochemical insights into natural variation in rice metabolism," *Nat. Genet.*, 2014, vol. 46, p. 714–721.

- [192] Y. Yu, T. Tang, Q. Qian, Y. Wang, M. Yan, D. Zeng, B. Han, C. I. Wu, S. Shi, and J. Li, "Independent losses of function in a polyphenol oxidase in rice: differentiation in grain discoloration between subspecies and the role of positive selection under domestication," *Plant Cell*, 2008, vol. 20, p. 2946–2959.
- [193] H. Kuriyama and M. Kudo, "Complementary genes Ph and Bh controlling ripening black coloration of rice hulls and their geographical distribution," *Jpn. J. Breed.*, 1967, vol. 17, p. 13–19.
- [194] M. Maekawa, "Geographical distribution of the genes for black hull coloration," *Rice Genet. Newsl*, 1984, vol. 1, p. 104–105.